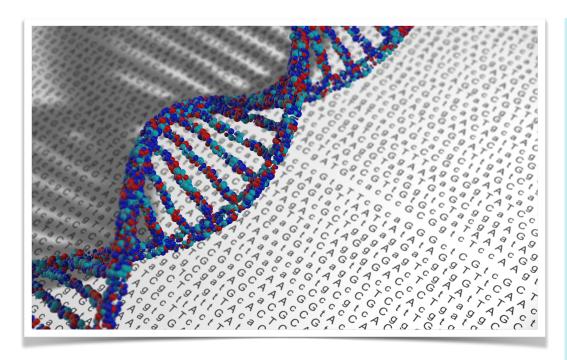


# Application note: NAPseq<sup>TM</sup>

Nucleic Acid Preservation Buffer, for NGS



## Suggested Volume of $NAPseq^{TM}$ Buffer

Briefly, the solid samples (such as cells, tissues, or feces) should be submerged in the buffer whereas the volume of liquid samples (such as saliva or biological fluids) should be equal to the volume of the NAPseq<sup>™</sup> buffer. The appropriate volume of NAPseq<sup>™</sup> buffer should be adjusted according to the samples as summarized in the table below.

/// Cell pellet : 1 ml for  $10^6$  cells / 2 ml for  $10^7$  cells

/// Tissue, Feces: 1 ml for 30 mg / 2 ml for 60 mg

/// Saliva : 1 : 1 ratio (for example 1 ml saliva : 1 ml  $NAPseq^{TM}$ )



### Storage

Reagent stable at room temperatures for at least 6 months.



### Sample Preservation

Stable at 25°C - 40°C Up to 7 days

# FREE SAMPLE

#### **EMAIL**

hi@bioentist.com

#### LINE

@bioentist

#### **WEBSITE**

www.bioentist.com

## **INHIBIT MICROBE**

Inhibit growth of microorganisms in the samples

### **STABLE**

Ensures nucleic acids stability during sample transport for 7 days at ambient temperatures.

#### CONVENIENT

No cold-chain or dry-ice needed during transportations

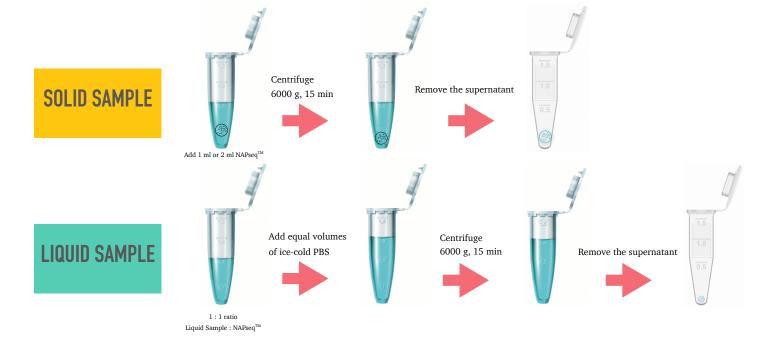
NAPSEQ<sup>™</sup> BIOENTIST CO., LTD.

## Sample Processing Before Nucleic Acid Extraction

For solid samples, centrifugation at 6000 g for 15 min to precipitate pellets and then remove the supernatant.

For biological fluid samples, add equal volumes of ice-cold phosphate buffered saline (PBS) before centrifugation at 6000 g for 15 min to precipitate pellets and then remove the supernatant.





## THE SAMPLE PELLETS ARE COMPATIBLE WITH SEVERAL METHODS FOR NUCLEIC ACIDS EXTRACTIONS

- 1. Magnetic-based Nucleic Acid Extraction Kits
- 2. Column based nucleic acids extraction kits
- 3. Phenol-Chloroform Extraction

